Rocaglamide breaks TRAIL-resistance in human multiple myeloma and acute T-cell leukemia in vivo in a mouse xenograft model

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A B S T R A C T
Multiple myeloma (MM) is an incurable malignancy by the presently known therapies. TRAIL is a promising anticancer agent that virtually not shows any toxicity to normal cells. We have recently carried out clinical trials with a human circularly permuted TRAIL, CPT, against MM saw a partial response in approximate 20–30% of patients. In the current study, we investigated the cause of CPT resistance and revealed that the majority of the MM patients express elevated levels of c-FLIP. Knockdown of c-FLIP expression by siRNA alone was sufficient to increase CPT-mediated apoptosis in a CPT-resistant human MM cell line U266. To overcome CPT resistance, we investigated the combination of CPT with Rocaglamide(s) in MM which has been shown to inhibit c-FLIP expression in vitro. We show that Rocaglamide(s) overcomes CPT resistance in U266 in vitro and significant increases in anti-tumor efficacies of CPT in mice xenografted with U266. Similar results were also obtained in mice xenografted with the CPT-resistant human acute T-cell leukemia cell line Molt-4. Our study suggests that the combination of Rocaglamide(s) with CPT may provide a more efficient treatment against myeloma and leukemia.

Introduction
Induction of apoptosis is one of the strategies against different types of cancer. So far, radio- and chemotherapy are the main treatments of cancer in addition to surgical resection of the tumor. However, toxicity to normal tissues is the main obstacle of radio- and chemotherapy. The tumor necrosis factor apoptosis inducing ligand (TRAIL) has emerged as one of the promising anticancer agents and chemotherapy. The tumor necrosis factor-induced death receptors TRAIL-R1/R2 (DR4/DR5) [6]. Comparison with the wild-type TRAIL, CPT shows improved stability and longer half-life. CPT also displays a better antitumor activity than TRAIL-R targeting agents and/or inhibitors of c-FLIP and IAPs [4].

Circularly permuted TRAIL (CPT) is a new generation of recombinant human TRAIL made by Beijing Sunbio Biotech LTD [6]. The primary difference of CPT from the wild-type TRAIL is that the N-terminus of amino acids 121–135 sequence is connected to the C-terminus of the amino acids 135–281 sequence of TRAIL by a flexible linker. This mutated molecule forms stable homotrimers and has potent apoptosis-inducing activity via enhanced binding to its death receptors TRAIL-R1/R2 (DR4/DR5) [6]. Comparison with the wild-type TRAIL, CPT shows improved stability and longer half-life. CPT also displays a better antitumor activity without significant toxicity against normal cells [6]. Recently, CPT has been studied in clinical trials in patients with relapsed and/or refractory multiple myeloma (RRMM) [7,8]. Multiple myeloma (MM) is a disease that affects plasma cells and can lead to devastating clinical features such as anemia, lytic bone lesions, hypercalcemia, and renal disease [9]. The clinical trials show that CPT mono-therapy is well tolerated but, unfortunately, only 20–30% of patients showed a partial or better response [7,8]. Currently, CPT is in a phase II clinical trial in
combination with the anti-angiogenesis drug Thalidomide in RRMM patients. It is thought that CPT and Thalidomide have different antitumor mechanisms and, thus, a combination of the two drugs would give a synergistic antitumor effect. However, the initial trial did not show the expected effect [10]. Therefore, it is necessary to find a rational CPT partner that can enhance TRAIL antitumor therapy in MM.

We have previously shown that a group of natural phytochemicals Cyclopenta[b]benzofuran compounds, collectively named Rocaglamides (Rocs) (also referred to as Flavaglines) (for reviews see Ref. [11] and Ref. [12]), selectively induce apoptosis in malignant cell by activation of the stress-response mitogen-activated protein kinases (MAPK) p38 and inhibit tumor growth by inhibition of the survival Raf-MEK-ERK signaling pathway leading to inactivation of eIF4E, a key factor for controlling the rate-limiting step of translation [13-15]. Due to translation inhibition, Rocs down-regulate expression of anti-apoptotic proteins which have a short half-life such as c-FLIP and Mcl-1 [14,16]. C-FLIP is a well-known inhibitor of the receptor-mediated signaling pathway and, thus, is the key determinant of tumor resistance to TRAIL-induced cell death [17,18]. Many cancers have been shown to over-express c-FLIP proteins including colorectal carcinoma, gastric carcinoma, pancreatic carcinoma, Hodgkin’s lymphoma, B cell chronic lymphocytic leukemia, melanoma, ovarian carcinoma, cervical carcinoma, bladder urothelial carcinoma, and prostate carcinoma [19].

We have recently shown that Rocs can sensitize resistant cancer cells towards TRAIL-induced cell death in HTLV-1-associated adult T-cell leukemia/lymphoma (ATL) and in Hodgkin’s lymphoma via inhibition of c-FLIP expression [14,20]. So far, the expression levels of c-FLIP in MM have not been thoroughly studied. Therefore, we asked whether CPT expression levels interfere with CPT therapy in MM and, if so, whether Rocs can overcome CPT-resistance in MM.

To answer the above questions, we investigated expression levels of c-FLIP proteins in primary MM samples isolated from patients. We show that the majority of MM patients express elevated levels of c-FLIP. We also show that over-expression of c-FLIP correlates with CPT resistance in MM cells. We further show that Rocs synergistically enhance CPT-induced apoptotic cell death in resistant MM cells in vitro. To confirm the in vitro data, we further carried out animal experiments and xenografted mice with either the CPT-resistant human MM cell line U266 or the CPT-resistant human T-cell leukemia cell line Molt-4, respectively. We show that combination treatment with Rocs and CPT significantly enhances efficacy of cancer treatment in vivo.

Materials and methods

Cell lines and cell culture

The human malignant cell lines used in this study are the T-cell acute lymphoblastic leukemias (T-ALL) Sup-T1, Molt-4 and Jurkat; the lymphoma T-cell line SeAx; the MM cell lines U266 and L363. All cells were cultured in RPMI 1640 glutamine (GIBCO) at 37 °C and 5% C and 5% C14 phosphate-buffered saline (PBS) several times. Bone marrow plasma cells were enriched by using the directly conjugated monoclonal mouse anti-CD138 microbeads (Miltenyi Biotec, Auburn, CA, USA) and by an immunomagnetic bead selection process as described previously [21]. Purity of cells more than 95% was confirmed by Z-color flow cytometry (CD138+ /CD45- and CD38+ /CD45- ) (Becton Dickinson, San Jose, CA, USA).

Preparation of human proliferating T cells

Primary T cells from human peripheral blood T cells were prepared to more than 90% CD3 positive as described previously [13]. For generation of proliferating T cells, freshly isolated T cells were cultured at 2 × 10^5 cells/ml and were activated with 1 μg/ml PHA overnight. Activated T cells were then washed three times and cultured for additional 5 days in the presence of 25 U/ml IL-2.

CPT and Rocs

CPT, the recombinant human mutant of TRAIL, is made by Beijing Sunbio Biotech LTD (Beijing, China), Roc-A (~98% pure) was purchased from Active Biochem, Hong Kong, China. The other Rocaglamide derivatives isolated from different species of the plant genus Aglaia are kindly provided by Dr. Peter Proksch (Institute of Pharmaceutical Biology and Biotechnology, University of Düsseldorf, D-40225 Düsseldorf, Germany). All compounds are >98% pure assessed by HPLC (more information see Ref. [15]). The synthetic Roc derivative FL3 is kindly provided by Dr. Laurent Désaube (Laboratoire d’Innovation Thérapeutique (UMR 7200), Faculté de Pharmacie, F-67401 Illkirch cedex, France).

Determination of apoptosis

Rocs were solved in dimethyl sulfoxide (DMSO); Roth, Karlsruhe, Germany) at a stock concentration of 50 mM. CPT was solved in cell culture medium. Cells were treated with different concentrations of Rocs or CPT for 48 h. Apoptotic cell death was examined by analysis of DNA fragmentation as previously described [13]. Results are presented as % specific DNA fragmentation using the formula: (percentage of experimental apoptosis - percentage of spontaneous apoptosis) / (100 - percentage of spontaneous apoptosis) × 100.

Knockdown of c-FLIP proteins

U266 cells were transfected with 200 nM of nonsense siRNA (#AM4611, thermo fisher scientific germany), C-FLIP, siRNA (5’-GAGCUUCUGAGACACCCUG-3’) and c-FLIP, siRNA (5’-CACCCUCAGGGCUAUGCGCU-3’) using the Amaxa Nucleofector solution (Nucleofector kit c) by the Amaxa program x-005 (Amlexa Biosystems, Cologne, Germany). After transfection, cells were allowed to rest for 24 h in culture and then treated with different concentrations of CPT for 24 h. After CPT treatment, cells were collected for apoptosis measurement. Knockdown efficiency of c-FLIP proteins was controlled by Western blot analysis 24, 48 and 72 h after transfection.

Western blot analysis

For each sample, 1 × 10^5 cells were lysed as previously described [13]. Equal amounts of protein were separated on 7-13% SDS-PAGE depending on the molecular sizes of the proteins and blotted onto a nitrocellulose membrane (Amschel Biosciences, Little Chalfon, UK) as previously described. The following antibodies were used: The Caspase-9 (sc-177784) from Santa Cruz, Dallas, Texas, USA; Caspase-3 (9662), XIAP (2042) and TRAIL-R2 from Cell Signaling Technology, Danvers, USA; TRAIL-R1 from Prosci, CA; tubulin and actin from Sigma, Saint Louis, USA; PARP (552428) from BD Biosciences, Heidelberg, Germany. The caspase-8 mAb C15 (mouse IgG2b) and c-FLIP mAb NF6 were generated in our lab as described previously [22,23].

Cell surface staining TRAIL receptors

For analysis of surface expression levels of TRAIL receptors, cells (5 × 10^5) were washed with PBS and incubated with 1 μg/ml of corresponding antibodies for 30 min at 4 °C, washed with PBS and incubated for 30 min with phycoerythrin-conjugated goat anti-mouse antibody (Dianova, Duabiva, Hamburg, Germany), and analyzed by flow cytometry with a FACSscan Cytometer (Becton Dickinson, Heidelberg, Germany). The following anti-TRAIL receptor antibodies were used: HS101 (TRAIL-R1/DR4), HS201 (TRAIL-R2/DR5), HS301 (DC-R1) and HS402 (DC-R2) (Enzo livesciences Lausen, Germany).

In vivo mouse studies

Immunodeficient mice (NOD CB17-Prkdcd scid/NcrCl) were implanted subcutaneously in the dorsal flank region with U266 (2.5 × 10^5) or Molt-4 (5 × 10^5) cells. In the U266 mouse experiment, once small tumors were visible (about two weeks after grafting), mice were randomly separated into four groups (8 mice in each group) and were treated by intraperitoneal injection (i.p.) with vehicle (DMSO) or with CPT (50 mg/kg) in the presence or absence of Roc-A (0.5 mg/kg) 5 times per week for 2 weeks and then further at 1 week on each second day. The tumor size was measured with a micrometer caliper and the tumor volume (V) was calculated by the formula V = (width × length) /2. In the Molt-4 mouse experiment, one week after grafting, tumor-size-matched mice (approximately 15 - 25 mm^3) were treated with vehicle or with CPT (50 mg/kg) in the presence or absence of Roc-A (0.5 mg/kg) (each group contains seven mice) five times per week by i.p. for two weeks. The mice were killed and the blood and tumor weight were measured. All protocols using and...
maintaining animals were approved by the German Animal Protection Authority (Office Regierungspräsidium Karlsruhe) in Karlsruhe.

Results

The majority of MM patient samples over-express c-FLIP proteins

As only 20–30% of MM patients show partial response to CPT, we decided to investigate the reasons that cause CPT resistance. We first selected two MM cell lines, a CPT-sensitive cell line L363 and a CPT-resistant cell line U266, as model systems (Fig. 1a). Western blot analysis showed that both cell lines express similar levels of Caspase-8 and Caspase-3 (Fig. 1b). Also, both cell lines express TRAIL receptors DR4 and DR5 (TRAIL-R1/R2) at the cell surface with a slightly higher level in L363 (Fig. 1c). No detectable expression of decoy receptor 1 (DcR1) and DcR2 were seen in both cell lines by Western blot analysis (data not shown) and by cell surface expression analysis (Supplementary Fig. S1). C-FLIP is the key inhibitor of the receptor-mediated signaling pathway [5,17,18]. Notably, U266 expresses significantly higher levels of c-FLIP, and c-FLIP\(_L\) compared to L363 (Fig. 1b). We than compared the c-FLIP expression levels of primary tumor samples from seven MM patients with L363 and U266. The analysis showed that 5 out of 7 MM samples (approximately 71% of the patients) express c-FLIP proteins at the levels similar to or even higher than U266 (Fig. 1d). XIAP is another inhibitor of the receptor-mediated signaling pathway [5]. Three patient samples show elevated XIAP expressions (about 43% of the patients) (Fig. 1d). The ratios of c-FLIP over-expressing patients consist with the CPT-response ratios in the clinical trials (20–30% of patients). Thus, high levels of c-FLIP expression are likely the cause of CPT resistance in MM patients.

Knockdown of c-FLIP increases CPT sensitivity in U266 cells

To investigate the role of c-FLIP in regulation of CPT sensitivity and resistance in MM cells, we performed a siRNA approach to knockdown c-FLIP expression in the CPT-resistant U266 cells. The experiment showed that specific knockdown of the expression of either c-FLIP\(_L\) or c-FLIP\(_S\) in U266 cells resulted in increase in CPT-mediated apoptotic cell death (Fig. 2). Despite the fact that U266 cells have less surface expression of DR4 and DR5 compared to L363 (Fig. 1c), down-regulation of c-FLIP protein expression is sufficient to enhance sensitivity of U266 towards CPT-induced killing. The experiment suggests that suppression of c-FLIP expression is a promising strategy to improve the TRAIL-based therapy in MM.

Rocs synergize with CPT to enhance apoptosis induction in U266 cells

We have previously shown that Rocs are potent suppressors of c-FLIP expression in HTLV-1-associated ATL and Hodgkin’s lymphoma cells [14,20]. Therefore, we asked whether Rocs could also overcome CPT-resistance in MM cells. To answer this question, we first tested the effect of Roc-A, the first Roc compound identified in 1982 [11,12], on U266 and L363. Cells were treated with an increasing concentrations of CPT in the absence or presence of different doses of Roc-A. U266 cells were highly resistant to CPT with only 10% cell death when treated with CPT alone even with a very high concentration (500 ng/ml), the concentration that killed almost 90% of L363 cells (Figs. 1a and 3a). Roc-A at the concentrations 10–40 nM induced also little cell death (10–20%). However, in combination of both drugs, Roc-A was shown to synergistically

![Fig. 1. Most of primary tumor cells from MM patients show elevated levels of c-FLIP.](image-url)

Cells were treated with different concentrations of CPT as indicated for 48 h. Apoptotic cell death was determined by DNA fragmentation. Means ± SD are shown. Results are representative of three independent experiments each performed in duplicate assays. b. Comparison of protein expression levels of caspase-8,-3, and c-FLIP in U266 and L363. Total cell lysates of U266 and L363 were subjected to Western blot analysis for expression levels of the proteins involved in receptor-mediated apoptosis. Actin was used for the loading control. Results are one of three independent experiments. c. Comparison of cell surface expression levels of the TRAIL receptors DR4 and DR5 in U266 and L363. Results are from one of two independent similar experiments. d. Analysis of c-FLIP and XIAP expression levels of MM patients samples. MM cells were isolated from seven patients and subjected to Western blot analysis for the expression levels of c-FLIP and XIAP. Actin was used as protein loading control.
enhance CPT-mediated cell death in a dose-dependent manner leading to almost 80% death in U266 cells (Fig. 3a). Synergistic effects were also observed in L363 cells treated by combinations of CPT and Roc-A (Fig. 3b).

To investigate whether other Rocs could also enhance the efficacy of CPT, we tested additional 11 Roc derivatives. All Rocs tested showed synergies with CPT to kill U266 cells (Fig. 3c). These experiments demonstrate that Rocs can enhance anticancer efficacy of CPT.

To investigate the mechanism by which Roc-A sensitizes CPT-mediated cell death, we carried out a kinetic analysis of the status of proteins involved in receptor- and mitochondria-mediated cell death (Fig. 4). U266 cells were treated with Roc-A and CPT alone or in combination for 4, 8, and 16 h. The cell lysates were subjected to Western blot analysis. As shown in Fig. 4, Roc-A (50 nM) alone was not able to activate pro-caspases and to induce PARP cleavage up to 16 h of treatment. Although treatment with CPT (50 ng/ml) alone slightly activated caspase-8, the amount of activated caspase-8 (p18) was too little to further activate pro-caspase-3 to generate the active p17 caspase-3. CPT alone was also not sufficient to induce PARP cleavage. In contrast, the combination of Roc-A and CPT resulted in strong activation of caspase-8 which in turn activated...
In vivo evaluation of the effect of CPT and Roc-A on MM

To explore the anti-tumor effect of combination therapy in vivo, we carried out an animal study. 2.5 × 10⁷ U266 cells each were implanted subcutaneously into immunodeficient mice. Two weeks after grafting, as small tumors were visible, mice were randomly separated into four groups (8 mice in each group) and were treated by intraperitoneal injection (i.p.) with vehicle (DMSO) or CPT (50 mg/kg) in the absence (DMSO) or presence of Roc-A (0.5 mg/kg) 5 times per week for 2 weeks and then further treated for 1 week on each second day. Tumor size was measured on days as indicated in Fig. 5a. The experiment showed that in the control group (DMSO), all 8 mice developed tumors. In the CPT treated group, 6 out of 8 mice showed delay in tumor growth for 26 days but developed tumors after stop of the treatment at the growth rate similar to the control group. In the Roc-A group, 4 mice (50%) were shown absence of tumor grow. In contrast, treatment with CPT in combination with Roc-A resulted in inhibition of tumor growth in 6 tumor baring mice (75%) (Fig. 5). This experiment demonstrates that combination of Roc-A with CPT significantly enhances efficacy of the therapy.

Discussion

So far, MM is a malignancy incurable by the presently known therapies. Despite of the development of several classes of new drugs, the median survival following treatment has remained at about 5 years for the past decade [24,25]. Thus, it is urgent to find new strategies for MM treatment. TRAIL is a promising anticancer agent which kills various cancer cells with no toxicity to normal tissues. So far, many efforts have been made to improve TRAIL-R-targeting agents. Although CPT and Roc-A alone are nontoxic to normal cells [6,13], the effect of combination of the two drugs on normal cells have not been examined. To investigate the tumor selectivity of the combination therapy, we examined the effect of CPT and Roc-A on proliferating normal blood T cells isolated from healthy donors. Consistent with previous data, no substantial cell death was seen by the combination of CPT and Roc-A in normal proliferating T cells (Supplementary Fig. S2). Thus, Roc-A does not enhance CPT toxicity in normal blood T cells.

To evaluate the effect of Roc-A and CPT on leukemia cells in vivo, 5 × 10⁶ of Molt-4 cells were grafted subcutaneously into mice. One week after xenografting, tumor-size-matched mice (approximatively 15 – 25 mm³) were treated with vehicle (DMSO) or with CPT (50 mg/kg) in the presence or absence of Roc-A (0.5 mg/kg) (each group contains 7 mice) five times per week by i.p. injection as described in Fig. 7a (upper panel). After two weeks of treatment, the body weight and tumor weight were measured. The experiment showed that CPT alone had almost no inhibitory effect on tumor growth (P = 0.529) (Fig. 7a and b). Treatment with Roc-A alone showed no significant reduction in tumor size but a substantial reduction in tumor weight (P = 0.040) (Fig. 7a and b). In contrast, the combination of both drugs significantly reduced tumor size as well as tumor weight (P = 0.005) (Fig. 7a and b). No body weight loss was seen in mice after treatment (Fig. 7c). The experiment demonstrates that Rocs can also improve the efficacy of TRAIL-based anticancer therapy in leukemia.

In vivo evaluation of the effect of CPT and Roc-A on leukemia

We further asked whether Roc-A could also improve TRAIL therapy in other types of hematopoietic malignancies. To investigate this question, we tested the effects of Roc-A and CPT on the human cutaneous T-cell lymphoma cell line SeAx and the human acute T-cell leukemia cell lines Jurkat, Sup-T1 and Molt-4. Treatment with CPT alone showed only 15–30% cell death in SeAx and Jurkat cells. Sup-T1 and Molt-4 cells were completely resistant to CPT treatment (Fig. 6a). Similar to MM cells, synergistic increases in apoptotic cell death were obtained by treatment with CPT in the presence of Roc-A in all four cell lines (Fig. 6a). Western blot analysis showed that down-regulation of c-FLIP expression correlated with increase in caspase-8 and 3 activities and enhanced cleavage of PARP (Fig. 6b). Thus, Roc-A can also enhance CPT-induced cell death in leukemia and lymphoma cells.

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We and others have previously shown that c-FLIP expression levels play an essential role in regulation of sensitivity and resistance of cancer cells towards TRAIL-mediated apoptotic cell death [14,17,18,20]. Previously, an analysis of c-FLIP expression levels in five different MM cell lines including U266 showed that all cells express comparable amounts of c-FLIP\(_L\) and low to undetectable levels of c-FLIP\(_S\). Based on this observation, the authors concluded that the levels of FLIPL were not predictive of the sensitivity of MM to TRAIL [35]. Therefore, we checked c-FLIP expression levels of primary MM cells and found that the majority of MM patient samples express elevated c-FLIP proteins (Fig. 1d). Knockdown of c-FLIP expression in the CPT-resistant U266 cells by c-FLIP specific siRNAs enhances CPT-mediated apoptosis (Fig. 2). This data indicates that c-FLIP also plays an important role in TRAIL resistance in MM cells.

Recently, we have shown in in vitro cell culture systems that the anti-cancer phytochemical Rocs can break TRAIL-resistance in HTV-1 ATL and Hodgkin's lymphoma cells through inhibition of c-FLIP expression [14,20]. Rocs are inhibitors of translation initiation [14,20,26]. Since c-FLIP proteins have a very short half-life [27,28], de novo inhibition of protein synthesis would consequently lead to down-regulation of c-FLIP expression and thereby enhance TRAIL-mediated cell death [14,20]. In this study, we extend our studies to MM and T-ALL. Consistent with our previous studies, the experiments show that Rocs synergize with CPT to enhance apoptotic cell death in MM and T-ALL cells (Figs. 3 and 6). These data suggest that
combinations of Rocs and CPT may be a rational strategy for cancer treatment.

So far, the effects of the combination of TRAIL and Rocs on cancer cells have not been tested in vivo. Therefore, we carried out animal experiments and xenografted mice with the human MM U266 and the human T-ALL Molt-4 cells, respectively. U266 and Molt-4 cells were resistant to CPT-mediated killing as shown in the apoptosis assays (Figs. 1a and 6a). Consistent with the in vitro data, the in vivo mouse experiments showed that treatment with CPT alone was not able to kill the tumor cells. The U266 tumor mouse experiment showed that CPT treatment for three weeks significantly delayed tumor growth. However, when treatment was stopped, tumors grew up with a similar rate as in the non-treated control group (Fig. 5b). This result may reflect the situation in CPT clinical trials. Treatment with CPT alone in the T-ALL Molt-4 mouse tumor model also showed no statistical significance in reduction of tumor weight (p = 0.529) (Fig. 7b). Natural Rocs have been shown to inhibit growth of several tumor cell lines in vivo in animal models including the mouse leukemia p388 [29], the mouse RAM lymphoma RMA [30], the human oral epidermis cancer KB [31], and the human breast cancer BC1 and MDA-MB-231 [32,33]. In our mouse experiments, treatment with Roc-A alone for two weeks in the T-ALL Molt-4 model showed a reduction in tumor weight (p = 0.040) (Fig. 7b). Also, treatment with Roc-A alone for three weeks resulted in tumor free mice (50%) grafted with U266 cells at the end of the experiment (10 weeks) (Fig. 5b). Significantly, treatment with a combination of CPT and Roc-A further reduced tumor weights (p = 0.005) in the T-ALL Molt-4 mouse model and in
the MM U266 mouse model 75% of mice were tumor free (Figs. 5b and 7b). Importantly, there were no influence in body weight on mice in the combination treatment in the T-ALL Molt-4 mouse model (p = 0.706) (Fig. 7c). There was also no loss of body weight in the MM U266 mouse model controlled on the day the mice were sacrificed (data not shown). Roc-A has no obvious toxicity up to a dose of 10 mg/kg body weight [32]. Acute toxicity of Roc-A was reported with an LD50 of >300 mg/kg [34]. The mouse experiments demonstrate that Rocs can be used to potentiate the anti-cancer activity of CPT and to increase the efficacy of cancer treatment.

In summary, we demonstrate that Rocs can break TRAIL resistance in MM and T-ALL in vivo. Our study raises the possibility to develop Rocs as a TRAIL adjuvant for the treatment of hematopoietic malignancies.

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Conflicts of interest

The authors have no conflicts of interest to disclose.

Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.canlet.2016.12.010.

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